

CHAPTER 12

THE USE OF ESSENTIAL OILS AGAINST SHEEP GASTROINTESTINAL NEMATODES

Filip Štrbac¹, Antonio Bosco², Ivan Pušić³, Dragica Stojanović¹, Nataša Simin⁴, Giuseppe Cringoli², Laura Rinaldi² and Radomir Ratajac³

¹Department of Veterinary Medicine, Faculty of Agriculture, University of Novi Sad, Serbia, Trg Dositeja Obradovića 8, 21102 Novi Sad, Vojvodina, Serbia

²Department of Veterinary Medicine and Animal Production, University of Naples Federico II, CREMOPAR, Via Federico Delpino I, 80137 Naples, Campania, Italy

³Scientific Veterinary Institute Novi Sad, Serbia, Rumenački put 20, 21113 Novi Sad, Vojvodina, Serbia

⁴Department of Chemistry, Biochemistry and Environmental Protection, Faculty of Sciences, University of Novi Sad, Novi Sad, Serbia, Trg Dositeja Obradovića 3, 21102 Novi Sad, Vojvodina, Serbia

*Corresponding author: strbac.filip@gmail.com

INTRODUCTION

Gastrointestinal nematodes (GINs) nowadays present a serious threat to sustainable sheep breeding. Various classes of commercial anthelmintics were used to control these parasites. However, due to the development of anthelmintic resistance (AR), the efficacy of these drugs has decreased, which in turn resulted in high economic losses. For these reasons, researchers are focused now on designing sustainable strategies for GIN control, based on the use of a combination of a bunch of options including the wise application of anthelmintic drugs, as well as applying alternative strategies. Within this context, phytotherapy (the use of plants or their products) presents one of the most promising alternatives. Essential oils (EOs) are natural, volatile and complex compounds characterized by a strong odor and extracted from aromatic plants. In various studies so far, these plant products showed high *in vitro* and, in some cases, *in vivo* efficacy against sheep GINs. The aim of this chapter is to review so far conducted studies based on the use of EOs against these parasites and to discuss results, as well as advantages of their use compared to commercial anthelmintics. On the other hand, current obstacles in the use of EOs and possible solutions on how to overcome them will be also discussed in this chapter. In this way, current and future perspectives of the use of EOs against sheep GINs are discussed here.

Sheep Gastrointestinal Nematodes and the Problem of Anthelmintic Resistance

Infections caused by gastrointestinal nematodes (GINs) are currently considered as one of the main obstacles for breeders of grazing sheep worldwide (Hammer et al. 2019). Although these infections are most commonly subclinical, manifested as impaired weight gain and lowered milk yields but in some cases they can lead to serious conditions such as anaemia, diarrhea, digestive problems, protein loss, lowered immunity and fertility and even death (Giovannelli et al. 2018; Bosco et al. 2020; Belecké et al. 2021). Therefore, the negative effect of these parasites is reflected in various ways, from impaired animal

health and welfare and reduced growth to a decrease in animal productivity and farm profitability (Velde et al. 2018). The economic losses caused by gastrointestinal parasitism are huge and difficult to estimate, although some reports indicate that these are estimated to be 17.94% of the total economic cost in animals (Abbas et al. 2020).

Nowadays, these parasites are widely distributed in many parts of the world. Generally, *Haemonchus* spp. and *Cooperia* spp. are more prevalent in sub-tropical/tropical environments, *Ostertagia* and *Nematodirus* spp. in the temperate regions, while *Trichostrongylus* spp. are prevalent throughout the world (Waller 2006). The prevalence of sheep GINs in Serbia is also high, with the following genera identified: *Nematodirus* spp. 71.22%, *Ostertagia* spp. 69.22%, *Trichostrongylus* spp. 66.55%, *Haemonchus* spp. 64.44% and *Chabertia* spp. 60.11% in Vojvodina, lowland landscape (Pavlović et al. 2017) as well as *Haemonchus* spp. (46.91%), *Oesophagostomum* spp. (40.73%), *Trichostrongylus* spp. (39.85%), *Nematodirus* spp. (35.88%) and *Chabertia* spp. (32.79%) in Eastern Serbia, predominantly mountainous (Kulišić et al. 2013). In southern Italy, the prevalence of sheep GIN genera varies but includes *Haemonchus* spp. (21-83%), *Trichostrongylus* spp. (2-59%), *Chabertia* spp. (0-48%), *Teladorsagia* spp. (0-25%) and *Cooperia* spp. (0-5%) (Bosco et al. 2020).

The control of sheep GINs is currently nearly exclusively reliant on commercial anthelmintic drugs (Bosco et al. 2020, Castagna et al. 2021). These include benzimidazoles (eg. albendazole, fenbendazole, mebendazole), macrocyclic lactones (eg. ivermectin, moxidectin, eprinomectin) and imidazothiazoles (eg. levamisole) (Dyary 2018; Velde et al. 2018). However, their improper use that refers to overfrequent treatments, miss-use or dose as well as continued use of one drug, has led to the development of anthelmintic resistance (AR) in different nematodes species and strains (Dyary 2018; Pinto et al. 2019; Bosco et al. 2020; Belecké et al. 2021), which is now reported worldwide. This has also been reported against even newly developed drugs such as monepantel (Mederos et al. 2014), whereby AR to a new drug has been reported in less than 10 years after introduction to the market. Furthermore, widespread incidence of multidrug-

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resistant populations of *Haemonchus contortus*, *Teladorsagia* and *Trichostrongylus* to benzimidazoles, imidazothiazoles and macrocyclic lactones in sheep throughout Europe has also been reported (Fissiha and Kinde 2021). Therefore, the reduced susceptibility of drugs in nematodes has resulted in even higher economic losses, which in turn endangers the sustainability of livestock (Vineer et al. 2020).

Sustainable Strategies for the Control of Sheep Gastrointestinal Nematodes

Modern sustainable strategies in sheep GINs management are based on rational use of anthelmintics along with the use of alternative strategies. Within the use of commercial anthelmintics, incorporation of refugia is suggested which is based on the treatment of only a proportion of animals instead of the whole group. In such conditions, one part of the parasite population remains untreated, which allows the maintenance of drug-sensitive parasites (Hodgkinson et al. 2019). The best-recommended practices for preserving refugia are targeted treatments (TT), that are related to the treatment of the whole flock based on knowledge of the risk or parameters that quantify the severity of infection as well as target selective treatments (TST), which are based on the treatment of only individual animals within the group to provide epidemiological and/or production benefits (Calvete et al. 2020). In essence, both TT and TST signify the rational use of drugs, i.e. on animals that require treatment due to health, productivity and welfare reasons, whereby single or combined pathophysiological and/or production-based treatment indicators are used for the selection of animals. These include clinical signs, faecal egg count (FEC), FAMACHA© scores, weight gain, milk production, body condition score (BCS), breech soiling and the diarrhoea score (DAG) (Soto-Barrientos et al. 2018; Calvete et al. 2020). Combining anthelmintics which have a related spectrum of activity but different modes of action, as well as the rotation of used anthelmintic classes, are also recommended as a way of slowing down the development of AR (Fissiha and Kinde 2021). On the other hand, these strategies could be complemented or even replaced with alternative solutions for controlling sheep GINs. Genetic control methods involve the selection of animals resistant to GIN, which as a process relies on the existence of genetic variation in the host and the predominant environmental conditions (Zvinorova et al. 2016). The objective of pasture management is to minimize the use of anthelmintics by avoiding exposure to parasite burden that would lead to clinical disease and loss of production, while simultaneously aiming to allow sheep to build up immunity (Abbott et al. 2012). Nutritional manipulation relies on a balanced grazing system that provides an adequate source of nutrients and an acceptable GIN burden, which in turn should allow an optimum level of productivity (Torres-Acosta et al. 2012). Biological control measures include the use of natural enemies against GINs, which mostly refers to different nematophagous fungal species such as *Duddingtonia* spp. (Szewc et al. 2021). In the end, the development of efficient vaccines against intestinal parasites will allow antiparasitic medications to be used less frequently (Fissiha and Kinde 2021).

Among the proposed alternative strategies for the control of sheep GINs, phytotherapy is considered particularly promising. Phytotherapy is defined as the use of plants to treat ailments, which as a healthcare practice is recorded since ancient times

and transferred from generation to generation (Borges and Borges 2016; Castagna et al. 2021). In veterinary medicine, there is an increasing interest in validating ethnoveterinary practices, judging by the high number of studies and articles on the topic (Eshetu et al. 2015). Antiparasitic properties are a common point of focus, whereby a wide range of plants or their products are considered suitable for the treatment of almost every parasitic disease in livestock (Athanasiadou et al. 2007). For this purpose, whole plants (e.g. rich in tannins), their parts or products such as extracts (e.g. aqueous and ethanol) and essential oils may be used. Within this context, plant based antiparasitic preparations may provide successful alternative remedies to synthetic drugs and be used in ethnoveterinary practices against sheep GINs (Castagna et al. 2021).

Properties and Chemical Composition of Essential Oils

Essential oils (EOs) are natural, volatile and complex mixtures of compounds characterized by a strong odor and extracted from aromatic plants (Bakkali 2008). As secondary metabolites, they are present in the specialized cells or glands and serve to protect plants from predators and pests, while also attract pollinators. These cells are present in all sections of these plants including flowers, leaves, buds, stems, twigs, seeds, fruits, roots, wood or bark (Bakkali 2008; Butnariu and Sarac 2018; Fokou et al. 2020). Therefore, EOs are considered as part of the plant immune system (Butnariu and Sarac 2018). As far as physical and chemical properties are concerned, EOs are liquid, volatile, and rarely colored, lipid-soluble and soluble in organic solvents with a generally lower density than that of water. These plant products are mostly extracted from flowers and leaves of various aromatic plants growing in temperate and warm regions such as the Mediterranean, the Amazon or tropical countries, where they represent an important part of traditional pharmacopoeia (Bakkali 2008).

The medicinal properties of EOs are recognized since ancient times which include antiviral, antibacterial, antifungal, antiparasitic, anti-inflammatory, antiseptic, anticancer and antispasmodic properties among others (Bakkali 2008; Mancianti and Ebani 2020; Zaman et al. 2020). In veterinary medicine, EOs are increasingly used for the prevention and treatment of various animal diseases. Although this still mostly refers to monogastric animals such as pigs and poultry (Mucha and Witkowska 2021), some implications and reports suggest their possible use as anthelmintic agents in ruminants as well (André et al. 2018). EOs may be obtained from plants by different methods of extraction, but the most common industrial methods are steam distillation and extraction with different solvents (Butnariu and Sarac 2018).

The active ingredients of EOs are responsible for their pharmaceutical effects. To date, approximately 5000-7000 different constituents of EOs are identified and described in which mono- and sesquiterpenes predominated, along with aromatic compounds such as phenylpropane derivatives (Morsy 2017; Butnariu and Sarac 2018) as mentioned in Table 1. Terpenes present polymers of isoprene (C₅H₈) and may be divided into hydrocarbons or oxygenated derivatives (oxides, alcohols, aldehydes, ketones, acids) or reaction products thereof (esters, ethers) (Butnariu and Sarac 2018). On the other hand, the group of phenylpropenes comprises constituents derived from n-propyl benzene (Morsy 2017). EOs have a very high variability of their composition, both in qualitative and quantitative terms (Dhifi et al. 2016).

Rational use of anthelmintics	Alternative strategies
<ul style="list-style-type: none"> •Refugia (TT and TST) •Combining anthelmintics •Rotation of used anthelmintic classes 	<ul style="list-style-type: none"> •Genetic resistance control •Pasture management •Nutrition adjustment •Biological regulation •Vaccine production •Phytotherapy (plants or their products such as extracts and essential oils)

Fig. 1: Strategies for sustainable control of sheep GINs.

Table 1: Constituents of essential oils and their division by chemical groups and examples (Dhifi et al. 2016; Morsy 2017; Butnariu and Sarac 2018)

Chemical group	Examples
Hydrocarbon terpenes	Limonene, α - and β -pinene, camphene, α - and γ -terpinene, sabinene, myrcene, β -caryophyllene, germacrene B and D, o and p-cymene etc.
Oxygenated derivatives (terpenoids)	a) Phenols - thymol, carvacrol b) Alcohols - linalool, citronellol, geraniol, menthol, α -terpineol, terpinen-4-ol, borneol c) Aldehydes - citral, citronellal, sinensal d) Ketones - α i β -tujon, camphor (2-bornenon), menton, carvone e) Oxydes - eucalyptol (1,8-cineole) f) Esters - linalyl-acetate, geraniol-acetate,
Phenylpropenes	Anethole, methyl chavicol (estragole), eugenol, vanillin, safrole, myristicin, cinnamaldehyde
Miscellaneous (sulfur- and nitrogen-containing compounds)	allyl sulfide, allicin, methyl anthranilate, indole, pyridine, pyrazine

Many factors such as light, precipitation, growing site (altitude, latitude), nature of the soil (pH, constituents), site of production and accumulation of the EOs in the plant, the age of the plant, the presence of soil organisms and microorganisms, predators and pollinators as well as postharvest treatment of EOs (Barra 2009; Fokou et al. 2020) may affect their chemical composition, which ultimately lead to variation in their pharmacological properties.

The Potential use of Essential Oils against Sheep Gastrointestinal Nematodes

In Vitro Tests

The interest of the use of EOs against sheep gastrointestinal nematodes, as well the number of studies upon that are in increasing trend over years. Within that context, different EOs showed anthelmintic potential so far. In vitro tests present the first step in the process of validating phytotherapy substances and are used for the initial evaluation and selection of plant species and their secondary metabolites that exhibit anthelmintic activity (Borges and Borges 2016; André et al. 2017; Štrbac et al. 2021a). Among in vitro tests, the most reliable and most common used tests are egg hatch assay (EHA) and larval development assay (LDA) that reflect ovicidal and larvicidal potential of EOs, as well as different larval and adult motility assays that suggest the effect of EOs on the motility of larva's or adults (Table 2). The advantages of the use of in vitro tests are ease of application, low cost, speedy, high reproducibility and no need for experimental animals

(protection of animal welfare), and thus have been widely used in the screening of medicinal plants, often rather than in vivo tests (Ferreira et al. 2016).

EOs listed in Table 2. showed anthelmintic potential against sheep GINs (mostly against *H. contortus*), but it differed depending on the oil used. The highest ovicidal activity, expressed as IC50 values, was recorded for *Cymbopogon schoenanthus* and *Cymbopogon martinii* (0.04 and 0.1 mg/mL, respectively, Katiki et al. 2011), *Thymus vulgaris* (0.098 mg/mL, Štrbac et al. 2021a), *Ruta chalepensis* (0.1 mg/mL, Akkari et al. 2015), and *Mentha arvensis* (0.1 mg/mL, Chagas et al. 2018). Different EOs of *Lippia* spp. showed great larvicidal activity with IC50 less than 0.01 mg/mL (Chagas et al. 2018), followed by *Thymus vulgaris* with IC50 as 0.062 mg/mL (Ferreira et al. 2016), *Hesperozygis myrtoides* with IC50 as 0.07 mg/mL (Castilho et al. 2017), *Piper aduncum* with IC50 as 0.1 mg/mL (Gáinza et al. 2016) and *Mentha piperita* with IC50 as 0.2 mg/mL (Katiki et al. 2011). *Cymbopogon schoenanthus* exhibited a very high activity on the larval motility with IC50 as 0.009 mg/mL (Katiki et al. 2012), whereby *Ruta chalepensis* induced 87.5% inhibition of motility of adults 8 h after exposure at the dose of 1 mg/mL (Akkari et al. 2015). Along with *Thymus vulgaris*, *Origanum vulgare*, *Foeniculum vulgare* and *Satureja montana* showed a high ovicidal effect in our studies with inhibition of egg hatchability up to 100% for each concentration tested, 0.049-50 mg/mL (Štrbac et al. 2021a; Štrbac et al. 2022). In certain experiments, bioactive compounds of EOs are also evaluated for anthelmintic activity, mostly against *H. contortus* and with the same tests. The list is also wide and includes anethole, B-elemene, borneol, camphor, carvacrol, carvone,

Table 2: Essential oils that have shown *in vitro* activity against sheep gastrointestinal nematodes, assays and references

Essential oil(s)	Assays	GIN species used	Reference
<i>Croton Zehneri</i> (two samples), <i>Lippia sidoides</i>	EHA, LDA	<i>Haemonchus contortus</i>	Camurça-Vasconcelos et al. 2007
<i>Eucalyptus globulus</i>	EHA, LDA	<i>H. contortus</i>	Macedo et al. 2009
<i>Cymbopogon schoenanthus</i> , <i>Cymbopogon martinii</i> , <i>Mentha piperita</i>	EHA, LDA, LFIA, LEA	<i>H. contortus</i> and <i>Trichostrongylus</i> spp	Katiki et al. 2011
<i>Arisaema lobatum</i> , <i>Arisaema franchetianum</i>	EHA, LDA, LMIA	<i>H. contortus</i>	Zhu et al. 2013a
<i>Artemisia lancea</i>	EHA, LDA, LMIA	<i>H. contortus</i>	Zhu et al. 2013b
<i>Tagetes minuta</i> , <i>Coriandrum sativum</i> <i>Alpinia zerumbet</i> , <i>Lantana camara</i>	EHA, LDA	<i>H. contortus</i>	Macedo et al. 2013
<i>Eucalyptus citriodora</i>	EHA, LDA	<i>H. contortus</i>	Ribeiro et al. 2014
<i>Melaleuca alternifolia</i>	EHA, LMIA	<i>H. contortus</i>	Grando et al. 2015
<i>Zanthoxylum simulans</i>	EHA, LDA, LMIA	<i>H. contortus</i>	Qi et al. 2015
<i>Cymbopogon citratus</i>	EHA, LDA	<i>H. contortus</i>	Macedo et al. 2015
<i>Ruta chalepensis</i>	EHA, AWMA	<i>H. contortus</i>	Akkari et al. 2015
<i>Citrus sinensis</i> , <i>Melaleuca quinquenervia</i>	EHA, LDA	<i>H. contortus</i>	Gaínza et al. 2015
<i>Thymus vulgaris</i>	EHA, LDA, LMIA, AWMA	<i>H. contortus</i>	Ferreira et al. 2016
<i>Piper aduncum</i>	EHA, LDA	<i>H. contortus</i>	Gaínza et al. 2016
<i>Hesperozygis myrtoides</i>	EHA, LDA	<i>H. contortus</i>	Castilho et al. 2017
<i>Lavandula officinalis</i> , <i>Citrus aurantifolia</i> , <i>Anthemis nobile</i>	EHA, LDA, AWMA	<i>H. contortus</i>	Ferreira et al. 2018
<i>Mentha arvensis</i> , <i>Zingiber officinale</i> , <i>Lippia sidoides</i> , <i>Lippia alba</i> , <i>Lippia origanoides</i> , <i>Lippia gracilis</i> , <i>Curcuma longa</i> , <i>Mentha piperita</i>	EHA, LDA	<i>H. contortus</i>	Chagas et al. 2018
<i>Rosmarinus officinalis</i>	EHA, LMIA	natural-mixed infection	Pinto et al. 2019
<i>Eucalyptus citriodora</i>	AWMA	<i>H. contortus</i>	de Araújo-Filho et al. 2019
<i>Origanum majorana</i>	EHA, AWMA	<i>H. contortus</i>	Abidi et al. 2020
<i>Juniperus communis</i>	EHA	natural-mixed infection: <i>Haemonchus</i> spp, <i>Trichostrongylus</i> spp, <i>Teladorsagia</i> spp and <i>Chabertia</i> spp	Štrbac et al. 2020a
<i>Coriandrum sativum</i>	LMIA	<i>H. contortus</i> , <i>Trichostrongylus axei</i> , <i>T. colubriformis</i> , <i>T. vitrinus</i> <i>Teladorsagia circumcincta</i> , and <i>Cooperia oncophora</i>	Helal et al. 2020
<i>Achillea millefolium</i> , two chemotypes	EHA	natural-mixed infection: <i>Haemonchus</i> spp, <i>Trichostrongylus</i> spp, <i>Teladorsagia</i> spp and <i>Chabertia</i> spp	Štrbac et al. 2020b
<i>Origanum vulgare</i> , <i>Satureja hortensis</i> , <i>Thymus vulgaris</i> , <i>Mentha piperita</i> , <i>Helichrysum arenarium</i>	EHA	natural-mixed infection: <i>Haemonchus</i> spp, <i>Trichostrongylus</i> spp, <i>Teladorsagia</i> spp and <i>Chabertia</i> spp	Štrbac et al. 2021a
<i>Cinnamomum verum</i> , <i>Syzygium aromaticum</i>	Mortality of nematode larvae in plant oil solution	<i>H. contortus</i>	Boyko and Brygadyrenko 2021
<i>Cinnamomum verum</i> , <i>Syzygium aromaticum</i> , <i>Melaleuca alternifolia</i> , <i>Piper cubeba</i> , <i>Citrus aurantiifolia</i> , <i>Lavandula angustifolia</i>	Mortality of nematode larvae in plant oil solution	<i>S. papillosus</i>	Boyko and Brygadyrenko 2021
<i>Ocimum basilicum</i> , 16 cultivares	EHA	<i>H. contortus</i>	Sousa et al. 2021
<i>Origanum vulgare</i> , <i>Pimenta dioica</i>	EHA, larval mobility	<i>H. contortus</i> and <i>Cooperia</i> spp.	Jiménez-Penago et al. 2021

*EHA - egg hatch assay; LDA - larval development assay; LMIA - larval motility inhibition assay; AWMA - adult worm motility assay, LFIA - larval feeding inhibition assay, LEA - larval exsheathment assay

citral, cinnamaldehyde, eucalyptol, eugenol, linalool, thymol, terpinen-4-ol and vanillin among the others (Katiki et al. 2017; André et al. 2018). In a study of Katiki et al. (2017), the highest ovicidal activity was shown by cinnamaldehyde, anethole, carvone, carvacrol and thymol with IC50 of 0.018, 0.07, 0.085, 0.11 and 0.13 mg/mL, respectively. The high larvicidal effect of carvacrol and thymol was demonstrated in studies of André et al. (2016) and Ferreira et al. (2016) with IC50 values of 0.2 and 0.06 mg/mL, respectively. The activity of these phenolic compounds may be associated with damage caused to the

cuticle and digestive apparatus on nematode larva's and neurotoxic effects on the free-living nematodes (interaction with SER-2 tyramine receptor) (André et al. 2016; 2017). Citral was also one of the most potent EO ingredients with an IC50 value of 0,13 mg/mL in EHA (Macedo et al. 2015). In some cases, the efficacy of binary, ternary and quaternary combination of EO isolated compounds was evaluated as well, whereby the highest ovicidal activities were shown by cinnamaldehyde:carvacrol (1:1), anethole:carvone (1:1) and anethole + carvone + cinnamaldehyde + carvone (1:1:1:1) with

Table 3: *In vivo* efficacy of essential oils against sheep gastrointestinal nematodes

Essential oil	Test, the time of evaluation and GIN species	Dose, routes of administration and duration of use	Efficacy	Reference
<i>Lippia sidoides</i>	FECRT; Days 0, 7, 14 and 21 a.t.	230 mg/Kg, oral, during 5 days 283 mg/Kg oral, during 5 days	38.0% at D7; 30.0% at D14; 29.8% at D21 45.9 at D7;-54.0% at D14; 22.9% at D21	Camurça-Vasconcelos et al. 2008
<i>Lippia sidoides</i>	Controlled test; Day 7 a.t.; <i>Haemonchus</i> spp. and <i>Trichostrongylus</i> spp.	283 mg/Kg, oral, during 5 days	<i>Haemonchus</i> spp. 56.9% <i>Trichostrongylus</i> spp. 39.3%	Camurça-Vasconcelos et al. 2008
Orange oil emulsion	FECRT; Days 0 and 14 a.t.; <i>H. contortus</i>	600 mg/Kg, single 600 mg/Kg during 3 days	97.4% 94.9%	Squires et al. 2010
<i>Cymbopogon schoenanthus</i>	FECRT; Days 0, 1, 5, 10, 15 and 20 a.t.; <i>H. contortus</i>	180 and 360 mg/Kg during 3 days, oral	n.e.	Katiki et al. 2012
<i>Cymbopogon schoenanthus</i>	Controlled test; Day 20 a.t.; <i>H. contortus</i>	180 and 360 mg/Kg during 3 days, oral	n.e.	Katiki et al. 2012
<i>Eucalyptus citriodora</i>	FECRT; Days 0,10 and 17 a.t.; <i>Haemonchus</i> spp., <i>Trichostrongylus</i> spp., <i>Oesophagostomum</i> spp.	250 mg/Kg	55.9% at day 10; 34.5% at day 17	Ribeiro et al. 2014
<i>Thymus vulgaris</i>	FECRT; Days 0, 2, 4, 6, 8, 10, 12 a.t.; <i>H. contortus</i>	75, 150 and 300 mg/Kg oral on Days 0, 6 and 12 a.t.	n.e.	Ferreira et al. 2016
<i>Mentha arvensis</i>	FECRT; Days 0, 1, 3, 7, 14 and 21 a.t.; <i>H. contortus</i> and <i>Trichostrongylus</i> spp.	200 mg/Kg, single dose	61.6% at D1; 48.1% at D14; 44.9% at D21	Chagas et al. 2018
<i>Cymbopogon citratus</i>	FECRT; Days 0, 8 and 15 a.t.; <i>Haemonchus</i> spp, <i>Trichostrongylus</i> spp and <i>Oesophagostomum</i> spp.	500 mg/Kg, oral, during 3 days	19.6% at D8; 23.9% at D15	Macedo et al. 2019
<i>Cymbopogon citratus</i>	Controlled test; Day 15 a.t. <i>H. contortus</i> , <i>T. colubriformis</i> , <i>O. columbianum</i> , <i>T. ovis</i>	500 mg/Kg, oral, during 3 days	<i>H. contortus</i> 66.4% <i>T. colubriformis</i> 38.4%	Macedo et al. 2019
<i>Eucalyptus citriodora</i>	FECRT, Days 0, 7 and 14 a.t. <i>Haemonchus</i> spp. <i>Trichostrongylus</i> spp., <i>Oesophagostomum</i> spp. and <i>Strongyloides</i> spp.	500 mg/Kg, oral, single dose	41.8% at D7; 69.5% at D14	de Araújo-Filho et al. 2019
<i>Thymus vulgaris</i>	FECRT; Days 0, 7 and 14 a.t.; <i>Haemonchus</i> spp.; <i>Trichostrongylus</i> spp.; <i>Teladorsagia</i> spp.; <i>Chabertia</i> spp.	100 mg/Kg, oral, single dose	25.23% at D7; 24.42% at D14	Štrbac et al. 2021b

* FECRT - faecal egg count reduction test; a.t. - after treatment; D - certain day after treatment; n.e. - not effective.

IC50 values of 0.012, 0.013 and 0.02 mg/mL, respectively (Katiki et al. 2017). In our study, the activity of linalool:estragole binary combination at a ratio 19%:81% exhibited ovicidal activity with IC50 of 0.98 mg/mL (Štrbac et al. 2021c). However, as many studies have demonstrated so far, an EO often shows higher anthelmintic activity in comparison with the single isolated compound, due to the synergistic effect among many different constituents of the whole EO, although it should be stressed that a wide number of compounds is not crucial for high efficacy (Štrbac et al. 2022).

In Vivo Tests

The results obtained through in vitro tests must be confirmed in field condition trials. For this purpose, various in vivo studies are used to obtain the most authentic results of the efficacy of plant-based formulations (Table 3). Although these studies can

be intensive, expensive and require time and animals for testing, these are essential as a further step in developing anthelmintic agents as they offer a clear picture of the possibility of using EOs and their ingredients against sheep GINs in everyday clinical practice. The most commonly used in vivo test is the faecal egg count reduction test (FECRT) which measures the percentage reduction in the number of nematode eggs excreted through faeces after administration of an active substance, and is confirmed by the controlled test that is based on the quantification of the worm burden after sacrificing animals which have previously been artificially inoculated with nematodes and treated (Kebede 2019).

As shown in Table 3, various EOs were found effective in different in vivo studies, whereby some were highly effective and some did not show any effect. Sometimes differences were found in efficacy of EOs of the same plant in different studies. This may be attributed to differences in chemical composition owing to variation in climate parameters, harvesting time, plant

parts used, solvents used for extraction etc. Thus, EO of *Thymus vulgaris* showed some anthelmintic effects in our study (Štrbac et al. 2021b), although it failed to reduce FEC of GINs in a study of Ferreira et al. (2016) at even greater doses. Those differences may be related to the different compositions and the isolate of EO used, which was confirmed in our study with *in vitro* tested *Achillea millefolium* EO (Štrbac et al. 2020b) or to the even other factors. The dependence of EO efficacy on the method of application (single or multiple uses) was also contradictory.

The *in vivo* efficacy of the isolated EO compounds or their combination was also evaluated in some cases. Some of them showed a high effect on the reduction of FEC, such as carvacrol-acetate and thymol-acetate with the efficacy of 65.9 % and 76.2 % on Day 14 a.t., respectively (doses of 250 mg/kg) (André et al. 2016; 2017). In a study of Chagas et al. (2018), pure menthol, at the dose of 160 mg/Kg, did not express *in vivo* efficacy unlike the whole oil whose main ingredient is, i.e. *Mentha arvensis* that reduced FEC by approximately 50% on Days 1, 7 and 14 at a similar dose tested, 200 mg/Kg. In our study, the efficacy of the binary combination of linalool: estragole (19:81%) in the FECRT at the single dose of 100 mg/kg was evaluated, whereby efficacy was found to be 24.21% and 25.90% on Days 7 and 14, respectively (Štrbac et al. 2021b).

Toxicity Studies

Rarely, toxicity studies of the use of EOs or their ingredients against sheep gastrointestinal nematodes have been conducted. In two studies on mice, LD50 values determined for carvacrol and thymol were 919 mg/Kg and 1350.9 mg/Kg, whereby for their acetylated derivatives carvacrol acetate (CA) and thymol acetate (TA), these values were 1544.4 mg/Kg and 4144.4 mg/Kg with no changes observed in the mice behavior (André et al. 2016; 2017). According to the guidelines proposed by Clark and Clarke (1977), orally administered substances with an LD50 value above 1000 mg/Kg are safe or considered as low-level toxic substances. So, CA, thymol and TA can be considered as non-toxic, while further studies should be performed for carvacrol. In a study of Ribeiro et al. (2014), the EO of *Eucalyptus citriodora* was classified as safe with an LD50 value of 2653.0 mg/Kg for mice. Some EO compounds such as menthol are of very low acute oral toxicity (LD50 > 2000 mg/Kg) (Chagas et al. 2018). Oral administration of the EO of *Origanum majorana* at different doses of 1000-5000 mg/Kg displayed no signs of toxicity, nor caused fatal effects in any of the treated mice during an observation period of 24 hours (Abidi et al. 2020). Katiki et al. (2012) concluded that *Cymbopogon schoenanthus* is safe for sheep at the doses of 180 mg/Kg and 360 mg/Kg, since no significant differences among group means for the hepatic (enzymes) or kidney (urea and creatinine) parameters were recorded after treatment with EO. In our *in vivo* studies, no toxic effects were observed on sheep, neither after oral administration of *Thymus vulgaris* (100 mg/mL) nor linalool:estragole (100 mg/mL) (Štrbac et al. 2021b).

Advantages and the Barriers of the use of Essential Oils to Control of Gastrointestinal Nematodes in Sheep

To date, EOs from various plants have shown efficacy against sheep GINs. As discussed above, their high anthelmintic potential is owed to various compounds that make up their composition, of which the primary component is most important (Dhifi et al. 2016). These compounds belong to

different chemical groups, which impart antiparasitic activity through different mechanisms of action and synergism. These involve interruption of the nematode nervous system, interference with the neuromodulator octopamine or GABA-gated chloride channels, the inhibition of AChE activity, disruption of the cell membrane of the nematode thereby changing its permeability, membrane and ion channel perturbations modifying membrane-bound protein activity and the intracellular signaling pathways inducing different neurological and structural changes leading to nematode paralysis and death (Andrés et al. 2012). Apart from the high efficacy, different chemical origins of their ingredients may contribute to less susceptibility of EOs to anthelmintic resistance (Macedo et al. 2010; Borges and Borges 2016). Moreover, the natural origin of plant-based formulations may contribute to less toxicity to hosts, as well as to fewer residues in meat and milk compared to synthetic drugs (Ferreira et al. 2018). Although this still needs to be confirmed, natural-based drugs are certainly much more environmentally acceptable. Finally, the use of chemical drugs is less and less sustainable not only due to AR, yet from the financial aspects, as drug prices continue to rise (Prakash et al. 2021). Also from that point of view, the use of different plant formulations could be a more sustainable and acceptable option given their low prices and ease of acquisition, especially in countries with developed biodiversity (Ferreira et al. 2018).

The main barriers in the use of EOs against sheep GINs as the widespread practice may be the lack of scientific data and trials aimed at verifying their efficacy against these parasites, which especially refers to *in vivo* trials. As discussed earlier, efficacy in field conditions must be proven before the use of some active substances in practice. Furthermore, toxicity studies should be conducted on the host animals. However, this field is relatively new and there is an increasing number of various studies aimed to confirm the efficacy and sustainable use of EOs against sheep GINs (Muthee 2018). Worsening of the situation due to AR forced many researchers worldwide to search for effective antiparasitic herbal formulations as a promising alternative to synthetic drugs. Our research group is actively engaged in the evaluation of new EOs for any anthelmintic efficacy through *in vitro*, *in vivo* and toxicity studies (data not shown).

The second problem about the current potential use of EOs in the practice is related to the low efficacy shown in field condition trials, which is still not comparable to commercially available anthelmintics (Macedo et al. 2010). Low *in vivo* efficacy is attributed to less bioavailability of active ingredients of EOs. This fact may be explained on the one hand by the anatomical and physiological specificity of the ruminant gastrointestinal tract (Hoste et al. 2008), and on the other hand by the unstable nature of EOs (Maes et al. 2019). Active ingredients of EOs are prone to evaporation and reaction with various factors inside the gastrointestinal tract. That leads to their partial or total inactivation before reaching the target place in abomasum or intestine. Thus, they usually show lower anthelmintic activity compared to that showed in different *in vitro* studies. Keeping in mind these factors, finding of plant species, the dose and route of administration effective *in vivo* is a challenge for ethnobotanists to be addressed. However, it also seems that increasing interest and the number of studies within this topic can contribute to overcoming this problem. Nevertheless, so far showed efficacy in different *in vivo* studies suggest that EOs and their active ingredients may be used as a valuable additional source in a nematode control along with

other measures, if not capable to be used independently (Macedo et al. 2010).

Encapsulation as a Novel Approach

Encapsulation is the method of protecting active components via physical or chemical processes. In this way, the active substance is physically separated from the environment by the creation of a protective shell, often referred to as the active carrier component or matrix (Lević et al. 2014). Nowadays different encapsulation techniques are used such as emulsification, nanoprecipitation and coacervation with chitosan, alginate or cyclodextrin as matrices (Maes et al. 2019). Given the instability and volatility of EOs, encapsulation could be of great importance when it comes to their application against GINs in sheep. Encapsulation reduces the interaction of the active substances with various factors in the environment (Radunz et al. 2018) and leads to reduced inactivation of active ingredients of EOs in the animal which ultimately results in increased bioavailability. Also, encapsulation represents a sustainable and efficient approach to increasing physical stability and protection against evaporation, enabling longer retaining properties and shelf life of EOs (Majeed et al. 2015). The other advantages of encapsulation include the increase the ease of handling active substances, reduction of odor and unpleasant taste (may be important for oral administration), as well as controlled release of the active substance (Radunz et al. 2018).

In a study of Mesquita et al. (2013), emulsified EO of (italic) given orally to the sheep at the dose of 365 mg/Kg once, reduced the total number of nematodes in sheep gastrointestinal tract by 60.79%, which was better than ivermectin that reduced the number up to 48.70%. The reduction of abomasal nematodes was significantly higher in the group treated with EO (83.75% and 35.00%, respectively). Similarly, the nanoemulsion of the same oil at the dose of 250 mg/Kg once, reduced the FEC of GINs similarly to levamisole ($p > 0.05$) in 8 of 10 days observed (Ribeiro et al. 2017). The dose of 250 mg/Kg of the encapsulated formulation of anethole:carvone (10% each and 80% of lipid matrix) given in food to lambs for 45 days significantly reduced FEC at the days 43 and 45, whereby the effect was attributed to a decrease in the size of males and a decrease in the fecundity of female nematodes (Katiki et al. 2019). At the same time, the formulation did not affect liver or kidney function of the lambs. When compared to free EO, nanoencapsulated oil of *E. citriodora* showed higher ovicidal (0.5 compared to 1.3 mg/mL) and similar larvicidal (both 1.7 mg/mL) in vitro, but slightly lower in vivo effect measured through faecal egg count reduction test at the same dose of 250 mg/Kg (40.5% compared to 55.9% at Day 10 a.t.) (Ribeiro et al. 2014). However, nanoemulsion of *C. citratus* showed clearly higher in vivo effect in the reduction of FEC compared to free EO, given orally for three days at the doses 450 and 500 mg/Kg, respectively (51.7% to 19.6% at Day 8 a.t.), whereby at the same time exhibited lower toxicity (Macedo et al. 2019). In most of these studies, chitosan was used as a carrier. However, studies aimed to confirm the positive impact of encapsulation techniques for the use of EOs against sheep GINs are needed.

Conclusion

In the era of AR, novel strategies for sustainable control of GINs in sheep farms should be designed. The use of EOs as an

alternative method show great potential due to their high efficacy originating from rich chemical composition, their affordable price and easy acquisition, especially in countries with developed biodiversity. Along with this, EOs possess significantly less susceptibility to resistance and better host and environmental acceptability from the toxicity aspect in comparison with synthetic drugs. The major obstacles are reflected in the lack of trials conducted in field conditions, as well as still usually lower in vivo effects than commercial drugs. However, these obstacles may be overcome with an increasing number of field studies in different conditions, especially with EOs and their ingredients that showed great in vitro potential. Moreover, applying novel methods such as encapsulation offers an opportunity to protect active EO ingredients from degradation and inactivation and thus allow a higher in vivo efficacy. From all the above, the use of these plant products may significantly contribute to the sustainable control of sheep GINs in the near future.

REFERENCES

- Abbas RZ et al., 2020. Anthelmintic effects and toxicity analysis of herbal dewormer against the infection of *Haemonchus contortus* and *Fasciola hepatica* in goat. *Pakistan Veterinary Journal* 40: 455-460.
- Abbott KA et al., 2012. Sustainable worm control strategies for sheep, 4th Ed., A Technical Manual for Veterinary Surgeons and Advisers, SCOPS, UK; pp: 40.
- Abidi A et al., 2020. Chemical analyses and evaluation of the anthelmintic effects of *Origanum majorana* essential oil, in vitro and in vivo studies. *Veterinarni Medicina* 65: 495-505.
- Akkari H et al., 2015. Chemical composition, insecticidal and in vitro anthelmintic activities of *Ruta chalepensis* (Rutaceae) essential oil. *Industrial Crops and Products* 74: 745-751.
- André WPP et al., 2016. Comparative efficacy and toxic effects of carvacryl acetate and carvacrol on sheep gastrointestinal nematodes and mice. *Veterinary Parasitology* 218: 52-58.
- André WPP et al., 2017. Anthelmintic effect of thymol and thymol acetate on sheep gastrointestinal nematodes and their toxicity in mice. *Revista Brasileira de Parasitologia Veterinaria* 26: 323-330.
- André WPP et al., 2018. Essential oils and their bioactive compounds in the control of gastrointestinal nematodes of small ruminants. *Acta Scientiae Veterinariae* 46: 1522.
- Andrés MF et al., 2012. Nematicidal activity of essential oils: a review. *Phytochemistry Reviews* 11: 371-390.
- Athanasiadou S et al., 2007. Medicinal plants for helminth parasite control: facts and fiction. *Animal* 1: 1392-1400.
- Bakkali F et al., 2008. Biological effects of essential oils – A review. *Food and Chemical Toxicology* 46: 446-475.
- Barra A, 2009. Factors affecting chemical variability of essential oils: a review of recent developments. *Natural Products Communications* 4: 1147-1154.
- Belecké A et al., 2021. Anthelmintic resistance in small ruminants in the Nordic-Baltic region. *Acta Veterinaria Scandinavica* 63: 18.
- Borges DGL and Borges FA, 2016. Plants and their medicinal potential for controlling gastrointestinal nematodes in ruminants. *Nematoda* 3: e92016.
- Bosco A et al., 2020. The threat of reduced efficacy of anthelmintics against gastrointestinal nematodes in sheep from an area considered anthelmintic resistance-free. *Parasites & Vectors* 13: 457.

- Boyko OO and Brygadyrenko VV, 2021. Nematicidal activity of aqueous tinctures of plants against larvae of the nematode *Strongyloides papillosus*. *Tropical Biomedicine* 38: 85-93.
- Butnariu M and Sarac I, 2018. Essential oils from plants. *Journal of Biotechnology and Biomedical Science* 1: 35-43.
- Calvete C et al., 2020. Assessment of targeted selective treatment criteria to control subclinical gastrointestinal nematode infections on sheep farms. *Veterinary Parasitology* 277: 109018.
- Camurça-Vasconcelos ALF et al., 2007. Anthelmintic activity of *Croton zehntneri* and *Lippia sidoides* essential oils. *Veterinary Parasitology* 148: 288-294.
- Camurça-Vasconcelos ALF et al., 2008. Anthelmintic activity of *Lippia sidoides* essential oil on sheep gastrointestinal nematodes. *Veterinary Parasitology* 154: 167-170.
- Castagna F et al., 2021. Green veterinary pharmacology applied to parasite control: evaluation of *Punica granatum*, *Artemisia campestris*, *Salix caprea* aqueous macerates against gastrointestinal nematodes of sheep. *Veterinary Sciences* 8: 237.
- Castilho CVV et al., 2017. In vitro activity of the essential oil from *Hesperozygis myrtoides* on *Rhipicephalus (Boophilus) microplus* and *Haemonchus contortus*. *Revista Brasileira de Farmacognosia* 27: 70-76.
- Chagas ACC et al., 2018. Efficacy of essential oils from plants cultivated in the Amazonian Biome against gastrointestinal nematodes in sheep. *Journal of Parasitic Diseases* 42: 357-364.
- Clark EGC and Clarke ML, 1977. *Veterinary Toxicology*, Cassel and Collier Macmillan, Publishers London, UK, pp: 268-277.
- de Araújo-Filho JV et al., 2019. Anthelmintic activity of *Eucalyptus citriodora* essential oil and its major component, citronellal, on sheep gastrointestinal nematodes. *Revista Brasileira de Parasitologia Veterinaria* 28: 644-651.
- Dhifi W et al., 2016. Essential oils' chemical characterization and investigation of some biological activities: a critical review. *Medicines (Basel)* 3: 25.
- Dyary HO, 2018. Anthelmintic resistance of gastrointestinal nematodes in sheep in Piramagroon sub-district, Sulaymaniyah/Iraq. *Tropical Biomedicine* 35: 373-382.
- Eshetu GR et al., 2015. Ethnoveterinary medicinal plants: preparation and application methods by traditional healers in selected districts of southern Ethiopia. *Veterinary World* 8: 674-684.
- Fissaha W and Kinde MZ, 2021. Anthelmintic resistance and its mechanism: a review. *Infection and Drug Resistance* 14: 5403-5410.
- Fokou JBH et al., 2020. Essential oil's chemical composition and pharmacological properties. In: El-Shemy H, editor. *Essential oils - oils of nature*. Intechopen, London, UK; chapter no. 2
- Ferreira LE et al., 2016. *Thymus vulgaris* L. essential oil and its main component thymol: Anthelmintic effects against *Haemonchus contortus* from sheep. *Veterinary Parasitology* 288: 70-76.
- Ferreira LE et al., 2018. Essential oils of *Citrus aurantifolia*, *Anthemis nobile* and *Lavandula officinalis*: in vitro anthelmintic activities against *Haemonchus contortus*. *Parasites & Vectors* 11: 269.
- Gaínza YA et al., 2015. Anthelmintic activity in vitro of *Citrus sinensis* and *Melaleuca quinquenervia* essential oil from Cuba on *Haemonchus contortus*. *Industrial Crops and Products* 76: 647-652.
- Gaínza YA et al., 2016. *Piper aduncum* against *Haemonchus contortus* isolates: cross resistance and the research of natural bioactive compounds. *Revista Brasileira de Parasitologia Veterinária*, 25: 383-393.
- Grando TH et al., 2016. In vitro activity of essential oils of free and nanostructured *Melaleuca alternifolia* and of terpinen-4-ol on eggs and larvae of *Haemonchus contortus*. *Journal of Helminthology* 90: 377-382.
- Giovanelli F et al., 2018. In vitro anthelmintic activity of four plant-derived compounds against sheep gastrointestinal nematodes. *Veterinary Sciences* 5: 78.
- Hammer K et al., 2019. The dynamics of ovine gastrointestinal nematode infections within ewe and lamb cohorts on three Scottish sheep farms. *Preventive Veterinary Medicine* 171: 104752.
- Helal MA et al., 2020. Nematocidal effects of a coriander essential oil and five pure principles on the infective larvae of major ovine gastrointestinal nematodes in vitro. *Pathogens* 9: 740.
- Hodgkinson JE et al., 2019. Refugia and anthelmintic resistance: concepts and challenges. *International Journal for Parasitology: Drugs and Drug Resistance* 10: 51-57.
- Hoste H et al., 2008. Identification and validation of bioactive plants for the control of gastrointestinal nematodes in small ruminants. *Tropical Biomedicine* 25: 56-72.
- Jiménez-Penago G et al., 2021. In vitro anthelmintic activity of *Pimenta dioica* and *Origanum vulgare* essential oils on gastrointestinal nematodes from sheep and cattle. *Journal of Parasitic Diseases: Official Organ of the Indian Society for Parasitology* 45: 583-591.
- Katiki LM et al., 2011. Anthelmintic activity of *Cymbopogon martinii*, *Cymbopogon schoenanthus* and *Mentha piperita* essential oils evaluated in four different in vitro tests. *Veterinary Parasitology* 183: 103-108.
- Katiki LM et al., 2012. Evaluation of *Cymbopogon schoenanthus* essential oil in lambs experimentally infected with *Haemonchus contortus*. *Veterinary Parasitology* 186: 312-318.
- Katiki LM et al., 2017. Synergistic interaction of ten essential oils against *Haemonchus contortus* in vitro. *Veterinary Parasitology* 243: 47-51.
- Katiki LM et al., 2019. Evaluation of encapsulated anethole and carvone in lambs artificially- and naturally-infected with *Haemonchus contortus*. *Experimental Parasitology* 197: 36-52.
- Kebede A, 2019. Review on anthelmintic drug resistance nematodes and its methods of detection in Ethiopia. *Journal of Veterinary Medicine and Animal Sciences* 2: 1013.
- Kulišić Z et al., 2013. Prevalence and intensity of infection with gastrointestinal nematodes in eastern Serbia. *Acta Veterinaria (Beograd)* 63: 429-436.
- Lević S et al., 2014. Savremeni procesi inkapsulacije u tehnologiji hrane. *Hrana i Ishrana (Beograd)* 55: 7-12.
- Macedo ITF et al., 2009. Ovicidal and larvicidal activity in vitro of *Eucalyptus globulus* essential oils on *Haemonchus contortus*. *Revista Brasileira de Parasitologia Veterinária* 18: 62-66.
- Macedo ITF et al., 2010. Anthelmintic effect of *Eucalyptus staigeriana* essential oil against goat gastrointestinal nematodes. *Veterinary Parasitology* 173: 93-98.
- Macedo ITF et al., 2013. In vitro effects of *Coriandrum sativum*, *Tagetes minuta*, *Alpinia zerumbet* and *Lantana camara* essential oils on *Haemonchus contortus*. *Revista Brasileira*

- de Parasitologia Veterinaria 22: 463-469.
- Macedo ITF et al., 2015. Anthelmintic activity of *Cymbopogon citratus* against *Haemonchus contortus*. *Revista Brasileira de Parasitologia Veterinaria* 24: 268-275.
- Macedo ITF et al., 2019. Anthelmintic effect of *Cymbopogon citratus* essential oil and its nanoemulsion on sheep gastrointestinal nematodes. *Revista Brasileira de Parasitologia Veterinaria* 28: 522-527.
- Maes C et al., 2019. Encapsulation of essential oils for the development of biosourced pesticides with controlled release: A review. *Molecules* 24: 2539.
- Majeed H et al., 2015. Essential oil encapsulations: uses, procedures, and trends. *RSC Advances* 5: 58449-58463.
- Mancianti F and Ebani VV, 2020. Biological activity of essential oils. *Molecules*, 25: 678.
- Mederos et al., 2014. First report of monepantel *Haemonchus contortus* resistance on sheep farms in Uruguay. *Parasites & Vectors* 7: 598.
- Mesquita MDA et al., 2013. Anthelmintic activity of *Eucalyptus staigeriana* encapsulated oil on sheep gastrointestinal nematodes. *Parasitology Research* 112: 3161-3165.
- Morsy NFS, 2017. Chemical structure, quality indices and bioactivity of essential oil constituents. In: El-Shemy H, editor. *Active ingredients from aromatic and medicinal plants*. Intechopen, London, UK; chapter no. 11.
- Mucha W and Witkowska D, 2021. The applicability of essential oils in different stages of production of animal-based foods. *Molecules* 26: 3798.
- Muthee JK, 2018. Anthelmintic efficacy of selected medicinal plants against gastrointestinal nematodes in naturally infected sheep in Kenya. *The Journal of Phytopharmacology* 7: 111-115.
- Pavlović I et al., 2017. Biodiversity of helminths of sheep breed in Vojvodina (northern Serbia). *Bulletin of University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca* 74: 162-166.
- Pinto NB et al., 2019. Ovicidal and larvicidal potential of *Rosmarinus officinalis* to control gastrointestinal nematodes of sheep. *Revista Brasileira de Parasitologia Veterinaria* 28: 807-811.
- Prakash P et al., 2021. Documentation of commonly used ethnoveterinary medicines from wild plants of the high mountains in Shimla District, Himachal Pradesh, India. *Horticulturae* 7: 351.
- Radunz M et al., 2018. A mini-review on encapsulation of essential oils. *Journal of Analytical & Pharmaceutical Research* 7: 00205.
- Ribeiro JC et al., 2014. Efficacy of free and nanoencapsulated *Eucalyptus citriodora* essential oils on sheep gastrointestinal nematodes and toxicity for mice. *Veterinary Parasitology* 204: 243-248.
- Ribeiro WLC et al., 2017. The use of *Eucalyptus staigeriana* nanoemulsion for control of sheep haemonchosis. *Pesquisa Veterinária Brasileira* 37: 221-226.
- Qi H et al., 2015. In vitro *Zanthoxylum simulans* essential oil against *Haemonchus contortus*. *Veterinary Parasitology* 211: 223-227.
- Soto-Barrientos N et al., 2018. Comparing body condition score and FAMACHA© to identify hair-sheep ewes with high faecal egg counts of gastrointestinal nematodes in farms under hot tropical conditions. *Small Ruminant Research* 167: 92-99.
- Sousa AIP et al., 2021. Essential oils from *Ocimum basilicum* cultivars: analysis of their composition and determination of the effect of the major compounds on *Haemonchus contortus* eggs. *Journal of Helminthology* 95: e17.
- Squires JM et al., 2010. Efficacy of an orange oil emulsion as an anthelmintic against *Haemonchus contortus* in gerbils (*Meriones unguiculatus*) and in sheep. *Veterinary Parasitology* 172: 95-99.
- Szewc M et al., 2021. Biological methods for the control of gastrointestinal nematodes. *The Veterinary Journal* 268: 105602.
- Štrbac F et al., 2020a. In vitro ovicidal effect of common juniper (*Juniperus communis* L.) essential oil on sheep gastrointestinal nematodes. *Veterinarski Pregled* 1: 152-159.
- Štrbac F et al., 2020b. In vitro ovicidal activity of two chemotypes of yarrow (*Achillea millefolium* L.) essential oil against ovine gastrointestinal nematode eggs. *Archives of Veterinary Medicine* 13: 59-76.
- Štrbac F et al., 2021a. Ovicidal potential of five different essential oils to control gastrointestinal nematodes of sheep. *Pakistan Veterinary Journal* 41: 353-358.
- Štrbac F et al., 2021b. In vivo potential of thyme (*Thymus vulgaris* L.) essential oil and synergistic combination of linalool:estragole to control sheep gastrointestinal nematodes. *Proceedings of 28th International Conference of the World Association for the Advancement of Veterinary Parasitology (WAAVP)*, Dublin, Republic of Ireland; 19-22 July 2021, pp: 517.
- Štrbac F et al., 2021c. In vitro ovicidal activity of mixture of linalool and estragole against gastrointestinal nematodes of sheep. *Veterinarski Pregled*, 2: 49-59.
- Štrbac F et al., 2022. Anthelmintic properties of essential oils to control gastrointestinal nematodes in sheep - in vitro and in vivo studies. *Veterinary Sciences*, 9: 93.
- Torres-Acosta F et al., 2012. Nutritional manipulation of sheep and goats for the control of gastrointestinal nematodes under hot humid and subhumid tropical conditions. *Small Ruminant Research* 103: 28-40.
- Velde FV et al., 2018. Farmer behavior and gastrointestinal nematodes in ruminant livestock- Uptake of sustainable control approaches. *Frontiers in Veterinary Science* 5:255.
- Vineer HR et al., 2020. Increasing importance of anthelmintic resistance in European livestock: creation and meta-analysis of an open database. *Parasite* 27: 69.
- Waller PJ, 2006. Sustainable nematode parasite control strategies for ruminant livestock by grazing management and biological control. *Animal Feed Science and Technology* 126: 277-289.
- Zaman MA et al., 2020. In vitro experiments revealed the anthelmintic potential of herbal complex against *Haemonchus contortus*. *Pakistan Veterinary Journal* 40: 271-273.
- Zhu L et al., 2013a. Anthelmintic activity of *Arisaema franchetianum* and *Arisaema lobatum* essential oils against *Haemonchus contortus*. *Journal of Ethnopharmacology* 148: 311-316.
- Zhu L et al., 2013b. In vitro ovicidal and larvicidal activity of the essential oil of *Artemisia lancea* against *Haemonchus contortus* (Strongylida). *Veterinary Parasitology* 195: 112-117.
- Zvinorova PI et al., 2016. Breeding for resistance to gastrointestinal nematodes – the potential in low-input/output small ruminant production systems. *Veterinary Parasitology* 225: 19-28.