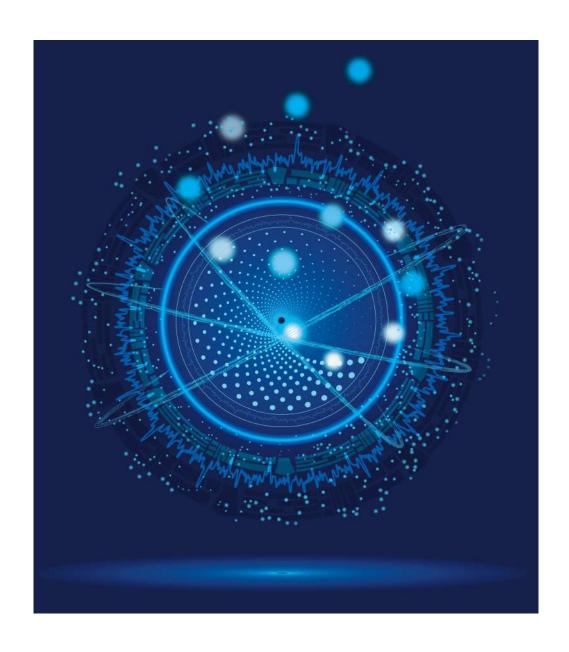
Small New World 2.0

4-5 September 2023

Abstract Book





Medical University Graz, Austria



Small New World 2.0

4-5 September 2023., Graz, Austria

Joint Meeting of









Austrian Society for Extracellular Vesicles - ASEV Hungarian Section for Extracellular Vesicles - HSEV Slovenian Network for Extracellular Vesicles - SiN-EV Serbian Society Extracellular Vesicles - SrbEVs

Organizing committee:

Beate Rinner, ASEV Wolf Holnthoner, ASEV Edit Buzas, HSEV Metka Lenassi, SiN-EV Maja Kosanović, SrbEVs

Scientific committee:

Beate Rinner, Medical University Graz, Austria;
Wolf Holnthoner, Ludwig Boltzmann Institute for Traumatology, Austria;
Edit Buzas, Semmelweis University, Hungary;
Metka Lenassi, Faculty of Medicine, University of Ljubljana, Slovenia;
Maja Kosanović, Institute for the Application of Nuclear Energy, INEP, Serbia;
Zoltan Giricz, Semmelweis University, Hungary;
Bernd Giebel, Institute for Transfusion Medicine, University Hospital Essen, Germany



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PROGRAM for Monday, 4th September 2023

8:30 - 10:00	Re	egistration and poster placement
10:00-10:15		om the Presidents of ASEV, HSEV, SiN-EV, SrbEV the local organizers & organizational introduction
10:15-12:00	EV therapeutics - regenerative medicine and beyond Chairs: Wolf Holnthoner (Austria) + Zala Jan (Slovenia)	
	Keynote: Bernd Giebel (Germany)	Clinical potential of MSC-EVs and translational challenges
	Beáta Szebeni	Origin of extracellular vesicles from peritoneal dialysate and their immunomodulatory effect
	Mateja Manček Keber	Therapy for cancer-specific MyD88L265P signaling based on exon skipping using LNP-mediated ASO delivery
	Katharina Schallmoser	A protein corona around human platelet-derived EVs promotes regenerative functions
	Maximilian Haertinger	Small extracellular vesicles derived from multipotent adipose stromal cells in peripheral nerve regeneration: jack of all trades, master of none?
	Alexander Otahal	Hoffa-derived MSCs primed with IL1β in bioreactor culture yields extracellular vesicles hindering chondrocyte recovery
12:00-13:30	0-13:30 Lunch break / General assembly of ASEV	
13:30-15:00	Methodology advances in EV analysis Chairs: Beate Rinner (Austria) + Sofija Glamočllija (Serbia)	
	Keynote: Pia Siljander (Finland)	Methods to study EVs – what do we (need to) look for?
	Jaroslaw Jacak	Tracking individual extracellular vesicles: from purification analysis to intracellular co-localization using atomic force- and single-molecule fluorescence microscopy
	Pietro Parisse	Substrate stiffness modulates extracellular vesicles' release in triple-negative breast cancer models
	Maria Jaritsch	Quantitative serum pharmacokinetics of EVs from diverse sources using high content single vesicle imaging
	Anna Lischnig	Quantitative proteomics and nFCM analysis of subpopulations of immune cell-derived EVs
15:00-15:30		Coffee break

PROGRAM for Monday, 4th September 2023

15:30-16:40	News from industry and development - "Rising projects" Chairs: Dirk Strunk (Austria) + Pia Siljander (Finland)	
	Clemens Helmbrecht ParticleMetrix	NTA goes colocalization: Characterization of Multi-labelled bionanoparticles
	Mehdi Madi and Quentin Lubart Abbelight	Quantitative analysis of single EV and their subpopulations with super-resolution solutions
	Core Facilities MedUni Graz	EV technologies at the MedUni Graz
	BioTechMed consortium "iNterAcD+"	Extracellular vesicle in exercise: sporty messengers in interorgan communication
	Christian Wadsack and Michaela Klaczynski	Fetal immune priming by placenta-derived small extracellular vesicles
	Beate Rinner and Mariangela Garofalo	Patient-derived tumor models, EVs and oncolytic viruses
16:40-16:45		Short break
16:45-17:30	Special guest lecture:	Translation of EV into the clinics - Eva Rohde (Austria)
17:30-23:00		
		Poster party and Social evening

PROGRAM for Tuesday, 5th September 2023

09:00-10:45	EV numbers and cargo Chairs: Maja Kosanović (Serbia) and Nicole Maeding (Austria)	
	Keynote: Paolo Bergese (Italy)	Extracellular vesicles by the numbers
	Hargita Hegyesi	Cardioprotective role of extracellular vesicle-mediated mir-sponge transfer
	Christa Noehammer	Small RNA biomarker profiling from extracellular vesicles in immune-mediated inflammatory diseases
	Tasvilla Sonallya	Systematic investigation and classification of membrane active peptides based on their affinity for interaction with extracellular vesicles
	Ilona Barbara Csordás	Extracellular Vesicles (EVs) miRNA-cargo loading and alterations after ionizing radiation induced cellular stress
	Marija Holcar	Characterization and Interindividual Variability of Plasma Extracellular Vesicles in Healthy Adults

10:45-11:30 Coffee break

11:30-12:30	NETWORK SESSION + MOVE Chairs: Beate Rinner and Wolf Holnthoner	
	Wolf Holnthoner	ASEV - Austrian Society for Extracellular Vesicles
	Edit Buzas/Zoltan Giricz	HSEV - Hungarian Society for Extracellular Vesicles
	Metka Lenassi	SiN-EV - Slovenian Network for Extracellular Vesicles
	Maja Kosanović	SrbEVs - Serbian Society for Extracellular Vesicles
	Johannes Oesterreicher	MOVE news from Finland
	Martin Wolf	M0VE news from Sweden

12:30-13:30 Lunch break

PROGRAM for Tuesday, 5th September 2023

13:30-15:00	Diversity of EV sources Chairs: Edit Buzas (Hungary) + Djenana Vejzovic (Austria)	
	Keynote:	Extracellular vesicle-mediated RNA delivery:
	Pieter Vader (The Netherlands)	from mechanistic insights towards therapeutic applications
	Astrid Laimer-Digruber	Unraveling the pathogenic and pro-inflammatory potential of extracellular vesicles secreted by Bacillus cereus
	Vendula Pospíchalová	Proteomic analysis of ascitic extracellular vesicles describes tumor microenvironment and predicts patient survival in ovarian cancer
	Kaja Ujčič	Effects of placental extracellular vesicles on maternal hematopoiesis
	Veronika Kralj-Iglič	Mechanisms of formation of extracellular particles in diverse samples from human, animal, plant and microalgae
15:00-15:30		Coffee break
15:30-17:00	Purity meets function Chairs: Metka Lenassi (Slovenia) + Krisztina Nemeth (Hungary)	
	Keynote:	

15:30-17:00	Purity meets function Chairs: Metka Lenassi (Slovenia) + Krisztina Nemeth (Hungary)	
	Keynote: Saara Laitinen (Finland)	To EV, or not to EV: that is the question
	Martin Wolf	Functional implications of protein EV corona
	Johannes Grillari	EV therapeutics - regenerative medicine and beyond
	Maria Cavinato	Alternative mechanisms of mitochondria quality control elicited by EVs in skin aging and disease
	Irma Schabussova	Outer membrane vesicles of the probiotic E. coli 083 activate innate immunity and prevent allergic airway inflammation in mice

17:00-17:15	Awards: Best poster & Best oral presentation Farewell notes
18:00	City tour Graz

Poster presentation P37



Surface-Associated Glycans as a Possible Distinct Factor for Establishing the Molecular Properties of Prostasomes

Jelena Danilović Luković; Tamara Janković; Sanja Goč, Filip Janjić, Ninoslav Mitić University of Belgrade, Institute for the Application of Nuclear Energy, INEP, Belgrade, Serbia

Glycans, complex carbohydrates bound to lipids and proteins, play important roles in biological processes. Prostasomes, extracellular vesicles from prostate epithelial cells, have a glycan composition influenced by their cellular origin and by the composition of the seminal plasma into which they are secreted. We hypothesized that the membrane-associated glycans could be used as selective targets for separation of prostasomes and as relevant parameter for distinction of their populations.

Prostasomes from seminal plasma of normozoospermic and oligozoospermic men were separated by ion-exchange- and lectin-affinity chromatography using concanavalin A (ConA) lectin, specific for mannosylated structures, and wheat germ agglutinin (WGA), specific for sialic acid. The presence of tetraspanins (CD63, CD9 and CD81), galectin-3 (gal-3) and gamma-glutamyl transferase (GGT) as known vesicle markers were monitored in association with distinct glycans. Their distribution was also analysed upon treatment of prostasomes with non-ionic detergent.

Membrane-associated GGT in the context of Con A- and WGA-reactive glycans mark prostasome populations from normozoospermic and oligozoospermic men. The assembly of tetraspanins, gal-3, and distinct N-glycans defines the solubilisation signature of prostasomes. WGA-reactive glycoproteins co-localize with CD9 and gal-3 in detergent-resistant domains, whereas ConA-reactive glycoproteins were distributed in detergent-sensitive domains along with CD63 and GGT. Subtle differences in the composition/presentation of examined molecules made difference among vesicles sharing the same physical properties in each group as well as between them.

The results obtained suggest the potential of glyco-parameters as reference markers for EVs populations.

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Disclamer: The authors are responsible for the contents of their abstracts and warrant that their abstract is original.

